

Finding *Schistocerca* genes in NCBI and making SnapGene files for cloning and alignments

What do you need?

1. NCBI website to find the genome and retrieve annotated GenBank files
<https://www.ncbi.nlm.nih.gov/>
2. FlyBase or other server to find gene orthologs you want to find in *Schistocerca*
<https://flybase.org/>
3. Some basic knowledge of BLAST
<https://blast.ncbi.nlm.nih.gov/Blast.cgi>
<https://chanzuckerberg.zendesk.com/hc/en-us/articles/360050963352-A-Guide-to-BLAST>
4. Gene visualization/manipulation software to look at/manipulate retrieved annotated Gene file

SnapGene which has a free version called SnapGene Viewer

<https://www.snapgene.com/snapgene-viewer>

Benchling

<https://www.benchling.com/molecular-biology>

What will you be able to do once you learn this tutorial?

With experience, it will take ~10-15 min to retrieve an annotated GenBank file of your favorite gene and open it in SnapGene to further work with for Protein sequence alignments and building trees for example

With an annotated sequence from one *Schistocerca* species, you will find and retrieve the orthologous sequences in the other *Schistocerca* species in 5-10 min

Note: Retrieving an annotated gene model from NCBI assumes that model is correct. All models are computer generated and may have errors. Once you do sequence comparison with the orthologous genes in different species errors may become apparent and these should be confirmed by the transcriptomic data. Monica's approach in Apollo will help verify and modify existing gene models.

Go to NCBI

Type in Schistocerca in the search bar

Go to the bottom left box, Genomes

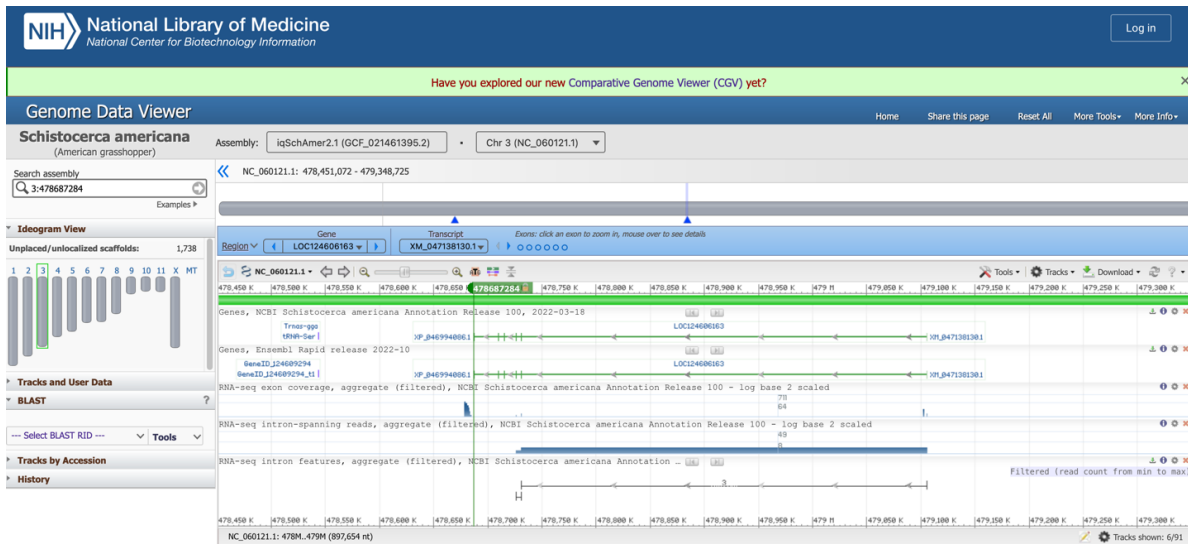
Go to first line, Assembly (there should be 7)

Click on the genome you want to search, for example *Schistocerca americana* (iqSchAmer2.1)

On the top right of the page, under Access the data, click on Genome Data Viewer

This brings you to the page below:

https://www.ncbi.nlm.nih.gov/genome/gdv/browser/genome/?id=GCF_021461395.2



There are 3 ways you can find a gene

1. Use a search term in “Search assembly”
2. Blast the CDS of the fly ortholog
3. Blast the protein sequence of the fly ortholog

First method: In search assembly, you can type in a gene name, for example “hyperpolarization” (this will find 4 genes that are of the HCN family)

Click on the one or ones that you want to download in SnapGene and use for further analysis
The screen will jump to the gene diagram that you want to focus on.

Proceed to Step 3 in the procedure below to retrieve your annotated GenBank gene file

Second method: perform a BLAST search using the Drosophila ortholog CDS

Step 1: retrieve the CDS sequence from FLYBASE

Go to FlyBase

Type in the gene name in the Flybase Search window (in our example the fly gene is *ih*)

This brings up a page with gene hits or takes you directly to the gene page

In the gene page, go to the 2nd box labeled Genome Location

Under sequence tab you can click on the drop down menu with Gene Region

Select CDS and then click get sequence

This brings you to the page of the coding sequence

Copy the complete CDS of the *ih* gene

Step 2: Perform a BLAST search from within the Genome Data Viewer for Schistocerca

Go back to the Genome Data Viewer of *Schistocerca* to perform your Blast search with the fly CDS

Go to the Blast button below the chromosome diagram window and click on tools

Click New BLAST

The default BLAST is a blastn for nucleotide sequence

Paste the Drosophila CDS in the window

Optimize the blast to the lowest similarity “somewhat similar sequences” (*see below)

Click BLAST, the page will refresh until the search is finished

The screenshot shows the BLAST interface from the National Library of Medicine. The search parameters are as follows:

- Job Title: Nucleotide Sequence
- RID: 2PRB8PEU01N
- Program: BLASTN
- Database: genomic/7009/GCF_021461395.2
- Query ID: Icl|Query_328333
- Description: None
- Molecule type: dna
- Query Length: 1857

The Filter Results section includes:

- Organism: (empty field)
- Percent Identity: (empty field) to (empty field)
- E value: (empty field) to (empty field)
- Query Coverage: (empty field) to (empty field)

The table of sequences producing significant alignments is as follows:

Description	Scientific Name	Max Score	Total Score	Query Cover	E value	Per. Ident	Acc. Len	Accession
Schistocerca americana isolate TAMUIC-IGC-003095 chromosome 5, isSchAmer2.1, whole genome shotgun...	Schistocerca a...	193	914	61%	2e-45	75.90%	778298166	NC_060123.1

This method returned 1 hit with a very good score on Chr 5 of the *Sa* genome (https://chanzuckerberg.zendesk.com/hc/en-us/articles/360050963352-A-Guide-to-BLAST) Click on the hit, and you will find that the CDS shows high homology (~75%) to 7 pieces, which represent exons of the gene in the *Schistocerca americana* genome Copy the coordinates of the gene in the first hit highlighted in blue below:

[Download](#) ▾ [GenBank](#) [Graphics](#) Sort by: E value ▾

Schistocerca americana isolate TAMUIC-IGC-003095 chromosome 5, iqSchAmer2.1, whole geno
 Sequence ID: [NC_060123.1](#) Length: 778298166 Number of Matches: 7

Range 1: [241908242 to 241908518](#) [GenBank](#) [Graphics](#) ▾ [Next Match](#) ▲ [Previous Match](#)

Score	Expect	Identities	Gaps	Strand
193 bits(213)	2e-45	211/278(76%)	2/278(0%)	Plus/Plus

Features: [potassium/sodium hyperpolarization-activated cyclic nucle...](#)
[potassium/sodium hyperpolarization-activated cyclic nucle...](#)

```

Query 1313      CAGGTGATATTATCATAAAGGAGGGTACGATCGGCTACTAAGATGTACTTCATACAGGAGG 1372
Sbjct 241908242 CAGGTGACATCATCATCAAGGAAGGAACCATTTGGTACCAAAATGTATTTATTCAAGAAG 241908301

Query 1373      GCGTGGTGGACATTGTCTATGGCAACGGCGAGGTTGCCACCTCACTTTTCG-GATGGGTCT 1431
Sbjct 241908302 GTATAGTTGACATTGTCTATGGCAAAATGGAGAAGTTGCTACAAG-CTTAAGTGATGGCTCT 241908368

Query 1432      TATTTGCGTGAGATCTGTCTGCTGACCAATGCGCTGTGTGCCAGCGTGCAGCCGAA 1491
Sbjct 241908361 TATTTGGGGAAATCTGTCTGCTGACGAATGCACGTCGTGTGCCAGTGTGAGAGCAGAA 241908428

Query 1492      ACCTATTGCAATCTATTCTCGTTGAGCGTGGATCATTTCAATTGCGTTCTGGATCAGTAT 1551
Sbjct 241908421 ACTTACTGTAATCTCTTTTCTTATCAGTGGAAACATTTCAATGTCGTTCTAGACCAGTAT 241908488

Query 1552      CCGCTGATGCGCAAGACCATGGAGACTGTGGCCGCCGA 1589
Sbjct 241908481 CCTTTAATGCGTCGCACCATGGAAAGCGTGGCAGCAGA 241908518
  
```

To locate the gene in the Genome Data Viewer, copy the coordinate into the Search Assembly box preceded by the chromosome number, so in this case type “5:241908242” and click the arrow.

The screen to the right will jump to that specific base in the gene. Click the zoom out button on the menu bar above the chromosome until you see the whole gene diagram.

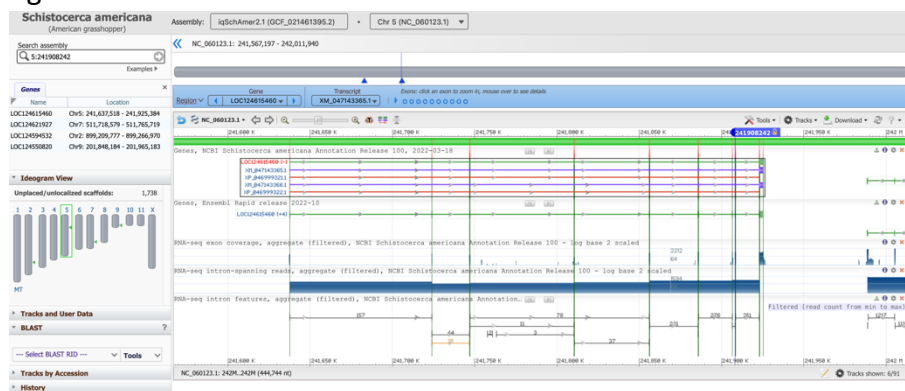
Step 3: Retrieve the GenBank file of your annotated gene

Click on the gene diagram

It will now add several colored lines to the diagram

1. Green represents the gene
2. Purple the mRNA
3. Red the protein

If you put your cursor on a line it will show a drop down menu with the information on the gene/mRNA/protein



On the green line (the zoomed out gene diagram) go down on the drop down menu to retrieve the GenBank file of the gene

It is the last line in the drop down menu: GenBank Record: NC_060123.1

This is the GenBank file of the gene LOC124615460

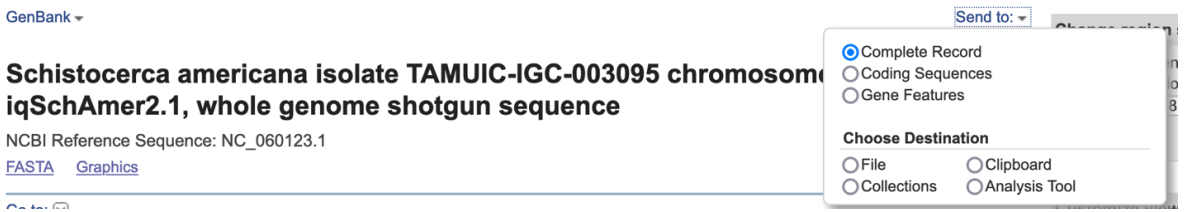
In this case this is one of 4 paralogs that encode a hyperpolarization cyclic nucleotide gated channel (this one is HCN2-like with the closest homology to *ih*)
 Scroll down and you will find all the information on the Gene under FEATURES

- Source
- Gene
- mRNA
- CDS, this file automatically includes the full translation of the gene

```

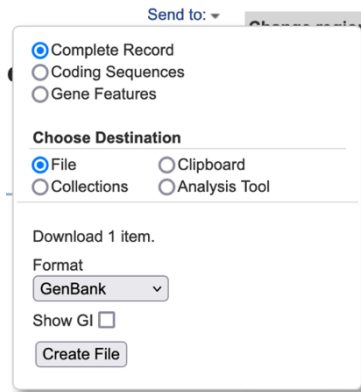
FEATURES             Location/Qualifiers
     source            1..287867
                        /organism="Schistocerca americana"
                        /mol_type="genomic DNA"
                        /isolate="TAMUIC-IGC-003095"
                        /isolation_source="physical"
                        /specimen_voucher="TAMUIC-IGC-003095"
                        /db_xref="taxon:7009"
                        /chromosome="5"
                        /sex="female"
                        /tissue_type="Whole body"
                        /dev_stage="adult"
                        /country="USA: Florida, St. Augustine"
                        /collection_date="2021-03-08"
                        /collected_by="Hojun Song"
                        /identified_by="Hojun Song"
     gene              1..287867
                        /gene="LOC124615460"
                        /note="Derived by automated computational analysis using
                        gene prediction method: Gnomon."
                        /db_xref="GeneID:124615460"
     mRNA              join(1..165,86453..86589,109518..109655,173035..173136,
                        218668..218817,248233..248451,248535..248681,
                        269165..269282,278728..271004,285443..287867)
                        /gene="LOC124615460"
                        /product="potassium/sodium hyperpolarization-activated
                        cyclic nucleotide-gated channel 2-like, transcript variant
                        X1"
                        /experiment="COORDINATES: polyA evidence [ECO:0006239]"
                        /transcript_id="XM_047143366.1"
                        /db_xref="GeneID:124615460"
     mRNA              join(1..165,86453..86589,109518..109655,218668..218817,
                        248233..248451,248535..248681,269165..269282,
                        278728..271004,285443..287867)
                        /gene="LOC124615460"
                        /product="potassium/sodium hyperpolarization-activated
                        cyclic nucleotide-gated channel 2-like, transcript variant
                        X2"
                        /experiment="COORDINATES: polyA evidence [ECO:0006239]"
                        /transcript_id="XM_047143366.1"
                        /db_xref="GeneID:124615460"
     CDS                1..165,86453..86589,109518..109655,173035..173136,
                        218668..218817,248233..248451,248535..248681,
                        269165..269282,278728..271004,285443..287867)
                        /gene="LOC124615460"
                        /note="Derived by automated computational analysis using
                        gene prediction method: Gnomon."
                        /codon_start=1
                        /product="potassium/sodium hyperpolarization-activated
                        cyclic nucleotide-gated channel 2-like isoform X1"
                        /protein_id="XP_046993321.1"
                        /db_xref="GeneID:124615460"
                        /translation="MILLLVNLLPLPAISFFNDLSTRMIAFNCLSDTIFLIDIVV
                        NFRGTIMQDQNAEQVLDPKLIAKHRLTWFFLDLSSIPLOYFLFNDFSEFQI
                        LMGRLRLRLAKLLSLVRLRLSRLVRYVSQWEVYVILNOKRRTERRGRSLDIA
                        PKKSKSKNLLFKFLNLSVFWLITLIDWLLIGHSGLQFLVPHLQFPFNSHW
                        AINELQGFNLEQYSNLFKMSHMLCIQYGRFPPOSITDMLTLMSISGATCYALF
                        LGHATNLIQSDSSRRYREKQVQVEEYMYRKLPREMRQRTIEYFHYQKGFDEE
                        AILGELSEKLREVDVNNKRSIVASVPPFNAQNSPVSQVVKLRVFPQDITIE
                        GTIGTNYFVQECIVDVMKGEVATSLSDSDFGECILLTNARVAVSVAETYNLF
                        SLSVEHFNVLDQYLPKRTMSVAERLNKIGNPNLVSHEEDMGSESKTINAVN
                        ALAEQAEHNTSEESVHGSSDKSIHELGRNLHELGLKTLHRLNPRKSENSFAASE
                        LPMRPAFHKSDFQKDTAFQ"
  
```

When you click on mRNA it will show all the annotated exons of the gene in the sequence file to the right
 To download this completely annotated gene file go to the top of the page to Send to and the drop down arrow
 It will show a box shown below:



Click on File, this will further drop down the menu and ask for a format

Default is GenBank
 Click on Create file
 This creates your annotated GenBank file
 When you open this file in SnapGene, you have a fully annotated Gene File



It is very useful to also download the mRNA and Protein and make a Snap Gene file of all the splice variants

Store all the labeled files in separate folders per gene in an Annotation Folder

These files will be useful to use to Blast the same gene in different *Schistocerca* species and to perform sequence alignments or build trees

*When using a *Schistocerca* CDS sequence to blast in another *Schistocerca* species genome to find the ortholog, make sure you use the best setting for the homology search.

Third method: *Drosophila* ortholog protein BLAST search

Instead of retrieving the CDS of the fly ortholog in Flybase retrieve the protein sequence

Go to FlyBase

Type in the gene name in the Flybase Search window

This brings up a page with gene hits or takes you directly to the gene page

In the gene page, go to the 2nd box labeled Genome Location

Under sequence you can click on the drop down menu with Gene Region

Select Translations and then click get sequence

This brings you to the page of the protein sequence

Copy the complete protein sequence

Next, go back to the Genome Data Viewer of *Schistocerca* to perform your Blast search with the fly protein sequence

Go to the Blast button below the chromosome diagram window and click on tools

Click New BLAST

The default BLAST is a blastn for nucleotide sequence

Switch to Tblastn before pasting the sequence

Paste the *Drosophila* protein sequence in the window

Click BLAST, the page will refresh until the search is finished

This method returned 3 hits, the top one is the same one as the CDS Blast search

The screenshot displays the BLAST search results interface from the National Library of Medicine. The top navigation bar includes the NIH logo and the text 'National Library of Medicine National Center for Biotechnology Information'. Below this, the BLAST logo and navigation links (Home, Recent Results, Saved Strategies, Help) are visible. The main content area is divided into several sections:

- Job Title:** Protein Sequence
- RID:** 2SR6PSV8013 (Search expires on 04-06 01:22 am, Download All)
- Program:** TBLASTN (Citation)
- Database:** genomic/7009/GCF_021461395.2 (See details)
- Query ID:** lcl|Query_36259
- Description:** unnamed protein product
- Molecule type:** amino acid
- Query Length:** 618
- Other reports:** (help icon)

The **Filter Results** section includes:

- Organism:** only top 20 will appear (exclude checkbox), with a text input field for 'Type common name, binomial, taxid or group name' and an '+ Add organism' link.
- Percent Identity:** [] to []
- E value:** [] to []
- Query Coverage:** [] to []
- Filter** and **Reset** buttons.

Below the filters, there are tabs for 'Descriptions', 'Graphic Summary', 'Alignments', and 'Taxonomy'. The 'Descriptions' tab is active, showing a section titled 'Sequences producing significant alignments'. This section includes a 'Download' dropdown, 'Select columns' dropdown, and a 'Show' dropdown set to '100'. A checkbox 'select all' is checked, indicating '3 sequences selected'. The table below lists the results:

	Description	Scientific Name	Max Score	Total Score	Query Cover	E value	Per. Ident	Acc. Len	Accession
<input checked="" type="checkbox"/>	Schistocerca americana isolate TAMUIC-IGC-003095 chromosome 5, IgSchAmer2.1, whole genome shotg...	Schistocerca a...	189	952	78%	5e-48	91.75%	778298166	NC_060123.1
<input checked="" type="checkbox"/>	Schistocerca americana isolate TAMUIC-IGC-003095 chromosome 9, IgSchAmer2.1, whole genome shotg...	Schistocerca a...	63.5	63.5	20%	8e-08	32.84%	242487659	NC_060127.1
<input checked="" type="checkbox"/>	Schistocerca americana isolate TAMUIC-IGC-003095 chromosome X, IgSchAmer2.1, whole genome shotg...	Schistocerca a...	57.4	57.4	9%	8e-06	45.90%	1012675668	NC_060130.1

There are 8 matches in the first hit with sequence identities of ~95%

The other 2 hits only have a single stretch that matches with much lower sequence identity (30-45%)

To complete the retrieval of any of the hits, proceed as in method 2 to complete the process.

Perform sequence alignments

What do you need?

1. Gene visualization/manipulation software to look at/manipulate retrieved annotated Gene file

SnapGene which has a free version called SnapGene Viewer

<https://www.snapgene.com/snapgene-viewer>

Benchling

<https://www.benchling.com/molecular-biology>

2. Clustal Omega online access

<https://www.ebi.ac.uk/Tools/msa/clustalo/>

SnapGene alignments

Go to SnapGene

Download all your protein sequences that you wish to align

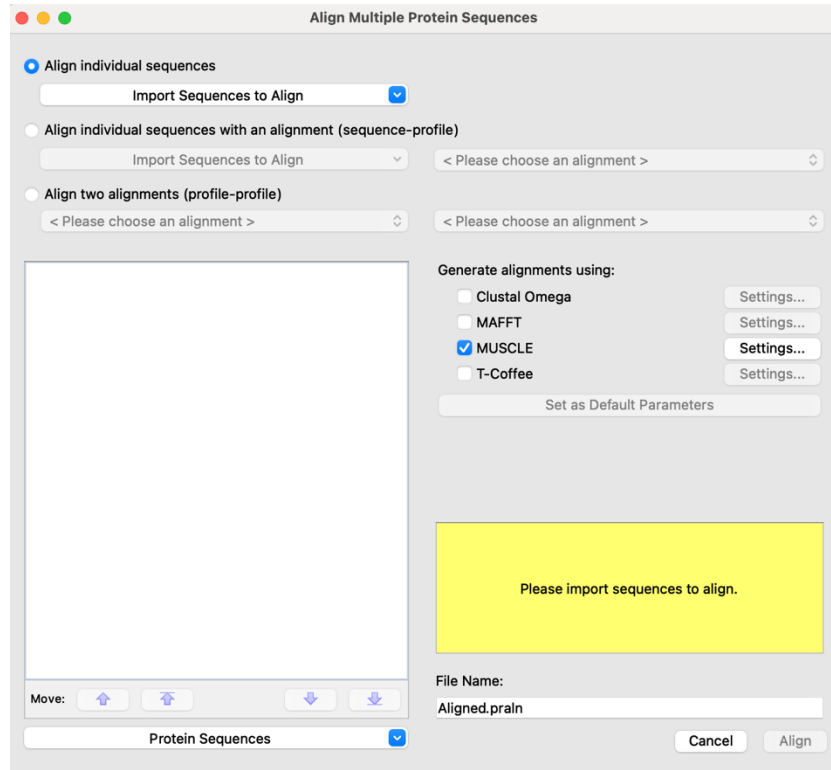
Make a folder to add all your protein sequence files with the names of the species and protein

Go to Tools

In the drop down menu go to Align Sequences

When aligning more than 2 sequences, choose Align Multiple Protein Sequences

This opens a box in which you can import all your sequences to be aligned



Import all the sequences from your folder

Select Clustal Omega and press align

This will show you an alignment of all the sequences

If you wish to omit a sequence, unclick it in the list of protein sequences

If you wish to change the output order, then use the up and down keys to move the sequences in the preferred order

Clustal Omega web alignments

Go to the Clustal Omega Website

Paste all your sequences you wish to align in the box

The easiest way to do this is to make a Word file with all your sequences

Make sure to start each sequence with > followed by a name and then paste the sequence on the next line

Repeat for all the sequences you wish to align

Multiple Sequence Alignment

Clustal Omega is a new multiple sequence alignment program that uses seeded guide trees and HMM profile-profile techniques to generate alignments between **three or more** sequences. For the alignment of two sequences please instead use our [pairwise sequence alignment tools](#).

Important note: This tool can align up to 4000 sequences or a maximum file size of 4 MB.

STEP 1 - Enter your input sequences

Enter or paste a set of

PROTEIN

sequences in any supported format:

Or, upload a file: No file selected. [Use a example sequence](#) | [Clear sequence](#) | [See more example inputs](#)

STEP 2 - Set your parameters

OUTPUT FORMAT

ClustalW with character counts

The default settings will fulfill the needs of most users.

(Click here, if you want to view or change the default settings.)

STEP 3 - Submit your job

Be notified by email *(Tick this box if you want to be notified by email when the results are available)*

If you use this service, please consider citing the following publication: [Search and sequence analysis tools services from EMBL-EBI in 2022](#)

Please read the provided Help & Documentation and FAQs before seeking help from our support staff. If you have any feedback or encountered any issues please let us know via [EMBL-EBI Support](#). If you plan to use these services during a course please contact us. Read our [Privacy Notice](#) if you are concerned with your privacy and how we handle personal information.

Use default parameters

Use default output format

When you submit, you will have the option to receive the results by email or just wait until the alignment is done

You can download the alignment file

You can also make a phylogenetic tree and download that file